

(GC-MS)
Salvia sclarea L. in vitro

Colletotrichum acutatum J.H. Simmonds

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Detailed GC-MS Analysis of the *Salvia sclarea* L. Essential Oil and the First *in vitro* Antifungal Activity Assessment against Crop Pathogen *Colletotrichum acutatum* J.H. Simmonds

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Original scientific paper

SUMMARY

<p><i>Salvia</i> L. (<i>Lamiaceae</i>) - 900 , - . <i>Salvia sclarea</i> L., - <i>Salvia</i>, , , (, , , -</p>	<p>The genus <i>Salvia</i> L. (<i>Lamiaceae</i>) comprises about 900 species widespread throughout the world. <i>Salvia sclarea</i> L., Clary Sage, one of the most appreciated representative of genus <i>Salvia</i>, is a biennial dicot shrub, native to southern Europe regions, but cultivated all over the world as a source of essential oil. Long history of use as a traditional medicine (antiseptic, anti-inflammatory, stomachic, digestive etc.) is further supported in the</p>
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(Foray et al., 1999; Pitarokili et al., 2002).

(Hudaib et al., 2001).

S.sclarea

S. sclarea.

MS

60

(43.17%),
(9.73%),
(1.43%).

(15.8%),
(5.13%)

GC GC-

D

S.sclarea,

Colletotrichum acutatum JH Simmonds C.A.2

, *in vitro*.

: *Salvia sclarea*,

, *Colletotrichum acutatum*

Salvia L.

Menthaeae

Nepetoideae

Lamiaceae 900

. *Salvia sclarea* L.,

Salvia,

numerous scientific studies (Foray et al., 1999; Pitarokili et al., 2002). Evaluation of secondary metabolism rate, modification in the qualities and quantities of essential oil constituents, and in particular the principal active ones affecting the biological activities is fundamental (Hudaib et al., 2001). The chemical composition of the essential oil of *S. sclarea* was found to be highly influenced by genetic and environmental factors, organ age, climate conditions, and seasonality. Herein, we present the results of detailed analyses of the essential oil constituents of the commercial sample of *S. sclarea* aerial parts. Plant material, harvested at full flowering stage from southeastern Serbian regions, yielded a transparent, yellowish fragrant essential oil. Subsequent meticulous GC and GC-MS analyses enabled the identification of more than 60 constituents, among which linalyl acetate (43.17%), linalool (15.8%), germacrene D (9.73%), caryophyllene (5.13%) and sclareol (1.43%) were the dominant ones. Together with the secondary metabolite profile, determination of the Serbian *S. sclarea* essential oil agricultural plant protection potential was estimated by assessing sporulation intensity and mycelia growth of *Colletotrichum acutatum* J.H. Simmonds C.A.2 isolates, causative of strawberry antrachnose, *in vitro*.

Key words: *Salvia sclarea*, Clary Sage, essential oil, antifungal, *Colletotrichum acutatum*

INTRODUCTION

Salvia L. is part of the tribe *Menthaeae* within the subfamily *Nepetoideae* of *Lamiaceae* with about 900 highly heterogeneous species. *Salvia sclarea* L., Clary Sage, is one of the commercial representatives in the genus *Salvia*, used in the perfumery industry, soft drink and liquor production, native to Mediterranean countries, North Africa and

(Werker et al., 1985).	central Asia, but nowadays cultivated worldwide (Werker et al., 1985).
<i>salvere</i> , " "	This species, with its name derived from the Latin <i>salvere</i> meaning to "heal", has a long history of use as a traditional medicine in the form of decoction or infusion with antiseptic, anti-inflammatory, stomachic, digestive and anticatarrhal properties (Foray et al., 1999; Pitarokili et al., 2002).
(Foray et al., 1999; Pitarokili et al., 2002). (EO), (Setzer, 2009), (Jirovetz et al., 2006; Kuzma et al., 2009), (Pitarokili et al., 2002; Fraternali et al., 2005; Jirovetz et al., 2007; Džami et al., 2008), (Özek et al., 2010), (Orhan et al., 2008), (Dikova, 2009) (Çınar et al., 2011).	Essential oil (EO) obtained from both wild and cultivated forms is proven effective through the numerous scientific studies in the treatment of anhyolitic effects (Setzer, 2009), antioxidant, antibacterial (Jirovetz et al., 2006; Kuzma et al., 2009), antifungal (Pitarokili et al., 2002; Fraternali et al., 2005; Jirovetz et al., 2007; Džami et al., 2008), antimalarial (Özek et al., 2010), anticholinesterase (Orhan et al., 2008), antiviral (Dikova, 2009) and opioid receptors activities (Çınar et al., 2011). The chemical composition of the essential oil of <i>S. sclarea</i> was found to be highly influenced by genetic and environmental factors, organ age, climate conditions, and seasonality. Evaluation of intra-species chemical polymorphism modification in the qualities and quantities of essential oil constituents, and in particular the principal active ones are of significance since, according to USP, BP and European pharmacopoeias, the quality control of the essential oils from medicinally important plants is the necessity.
<i>S. sclarea</i> BP	
<i>Colletotrichum acutatum</i> J.H. Simmonds (<i>Fragaria x ananassa</i> Duchesne, Rosaceae), (Freeman and Katan, 1997, Freeman et al., 2002).	<i>Colletotrichum acutatum</i> J.H. Simmonds is a plant pathogen infecting both the root and crown of strawberries (<i>Fragaria x ananassa</i> Duchesne, Rosaceae), causing necrosis manifested on fruits, stolons, leaf and flower stems (Freeman and Katan, 1997, Freeman et al., 2002). This fungus can attack the plant at all stages of development causing significant production losses worldwide, both before and after harvesting.

(Freeman and Shabi, 1996; Talhinhos et al., 2011). Since the first anthracnose, recorded in Europe in 1984 (Talhinhos et al., 2011), the disease has been present in Australia, New Zealand, United Kingdom the United States as well as in various countries in Asia, Africa and South America, for a long time. Yield reductions of over 80%, due to the occurrence of this pathogen, were observed in Serbia when the presence of *C. acutatum* on strawberries was first established (Ivanovi et al., 2005). Besides this main host, *C. acutatum* can also be present in other species important for agriculture, such as *Malus* sp., (Rosaceae), *Olea europaea* L., (Oleaceae), *Piper* sp. (Piperaceae), and many others indicating its overall pathogenic relevance (Lee et al., 2007; Peres et al., 2008; Polashock et al., 2009; Talhinhos et al., 2011). The control of the anthracnose causative agent, *C. acutatum*, is primarily aimed at applying available synthetic agents. However, the increasing requires for healthy food limits the use of synthetic pesticides and raises the need for new, efficient and safe, antifungal agents used against multi-resistant strains. Pesticides of herbal origin have received substantial attention since they represent a prolific source of a variety of bioactive compounds, with no or little harmful effect on non-target organisms and the environment. Furthermore, EOs constituents are identified as allelochemical, limiting the growth of competing plants in the surrounding environment, which is another advantage of their usage (Ulukanili et al., 2018). In the present study, we report on the results of the detailed GC-MS analyses of the *Salvia sclarea* L. essential oil constituents, originating from the southeastern Serbian regions, in order to determine qualitative and quantitative composition of the essential oil and evaluate the secondary

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Furthermore, EOs constituents are identified as allelochemical, limiting the growth of competing plants in the surrounding environment, which is another advantage of their usage (Ulukanili et al., 2018). In the present study, we report on the results of the detailed GC-MS analyses of the *Salvia sclarea* L. essential oil constituents, originating from the southeastern Serbian regions, in order to determine qualitative and quantitative composition of the essential oil and evaluate the secondary

C.A.2 *Colletotrichum*
acutatum J.H. Simmonds,
in vitro

metabolite rate in the sense of the modification in essential oil constituents.

Together with the secondary metabolite profile, determination of the agricultural plant protection potential of Clary Sage essential oil on *Colletotrichum acutatum* J.H. Simmonds C.A.2 isolates, causative of strawberry antrachnose, *in vitro*, is estimated by assessing sporulation intensity and mycelia growth.

MATERIAL AND METHODS

1. Plant material

A commercial sample of the essential oil of cultivated *Salvia sclarea* L. originating from Crvena reka, southeastern Serbia (latitude: 43°59'7.44", longitude: 19°57'20.41", altitude: 268 m) was utilized in this work.

1.

Salvia sclarea L.
 Crvena reka,
 (: 43°59'7.44",
 : 19°57'20.41",
 : 268 m).

2. GC-MS analysis

Mass spectra were recorded on a Hewlett-Packard 6890N gas chromatograph equipped with a fused silica capillary column DB-5MS (5% diphenyl, 95% dimethylpolysiloxane, 30 m × 0.25 mm, film thickness 0.25 μm, Agilent Technologies, Lexington, USA) and coupled with a 5975B mass selective detector from the same company. The injector and interface were operated at 250 °C and 300 °C, respectively. Oven temperature was raised from 70 to 290 °C at a heating rate of 5 °C/min; the heating program ended with an isothermal period of 10 min. As carrier gas helium, He, at 1.0 mL/min was used. The samples, 1 μl of the appropriate solutions in diethyl ether (1 mg/mL) were injected in a splitless mode. MS conditions were as follows: ionization voltage of 70 eV, acquisition mass range 35-650, scan time 0.32 s.

2. GC-MS
 Hewlett-Packard 6890N,
 DB-5MS (5%
 , 95%
 30 m × 0.25 mm,
 0.25 μm, Agilent Technologies, Lexington, USA)
 5975B
 250 °C 300 °C.
 70 290 °C
 5 °C/min;
 (He) 1.0 mL/min. , 1 μl
 (1 mg/ml),
 MS
 70 eV,
 35-650, 0.32 s.

3.

3. Identification of compounds

The constituents of the essential oils were identified by using their retention indices, calculated by linear interpolation

relative to retention times of a series of *n*-alkanes, and their mass spectra on a previously mentioned DB-5 column under the same chromatographic conditions.

Identification of individual compounds was made by comparison of their mass spectra with those of the internal reference mass spectra library, with the data bank mass spectra (Wiley 7N and NIST/NBS libraries) or with authentic compounds, and confirmed by comparison of their retention indices (using the generalized equation by Van Del Dool et al., 1963 and Adams, 2007) with authentic compounds or with previous literature reports. For quantification purpose, relative amounts of individual components were calculated based on GC-MS peak areas without FID response factor correction by assuming a unity response by all.

4. Antifungal activity

4.1. Test organism

In this experiment, the used isolate of *C. acutatum* J.H. Simmonds (C.A.2) was obtained from *Fragaria x ananassa* (Weston) Duchesne ex Rozier, Rosaceae grown in Serbia. The isolate was determined based on morphological, pathogenic and molecular characteristic, and maintained on a potato-dextrose agar (PDA) at the temperature of 25 °C.

4.2. Effect of volatile phase of essential oil on the growth of *Colletotrichum acutatum* isolates

The antifungal activity of essential oils was tested on PDA in Petri dishes 90 mm in diameter. Substrates were inoculated by sowing mycelial fragments of isolates taken from the edge of cultures aged seven days.

The oils were applied in the form of drops placed in the inside of the lid of the Petri dish in concentrations of 0.02, 0.04, 0.08

0.08 0.16 $\mu\text{L/ml}$

25 °C.

480 SC
(a.m. Captan, Galenika phytopharmacy)

480 SC

PDA,
(Grahovac, 2014).

30

(MIC).

(MLC) (Grahovac,
2014).

(ANOVA)
StatSoft

STATISTICA 8.0.

and 0.16 $\mu\text{L/mL}$ of air. Immediately after applying the oils, the Petri dishes were reversed and sealed with parafilm tape to prevent vapour loss.

Exposure of the isolates to the vapours of the studied oils lasted seven days at 25 °C. The assay was performed in four replicates for each oil concentration. The isolates grown on the medium with added fungicide Method 480 SC (a.m. Captan, Galenika phytopharmacy) were used as the positive control variant.

The Fungicide Method 480 SC was applied at a concentration recommended for practical application, diluted in sterile water and homogenized on a magnetic stirrer. Similarly, the negative control variant, fungi isolates grown on PDA were used under identical conditions without the described treatment (Grahovac, 2014).

Seven days after the treatment, the effect of essential oils was represented by the percentage of inhibition of mycelial growth, compared to the controls. The lowest concentration with the effect of complete inhibition of isolate growth was defined as the minimum inhibitory concentration (MIC). After the evaluation, Petri dishes, in which the growth of the isolate was completely stopped, were opened and ventilated under a stream of air in the laminar chamber for 30 minutes to remove the gaseous phase of the oil and determine the lethal effect.

It was considered that a certain concentration of oil exhibits a lethal effect if no initial growth of the isolate was noted seven days after ventilation. The lowest airborne effect concentration was defined as the minimum lethal concentration (MLC) (Grahovac, 2014).

The results obtained during the research were processed by ANOVA analysis with the statistical program StatSoft STATISTICA 8.0. Duncan's test was conducted to

P = 0.05

4.3.

C. acutatum.

Thom.

5 ml

C.A.2 (*C. acutatum*)

J.H. Simmonds). (Vasic,

2007).

Quesada

Lopez (1980), : +=

(<5.000 /ml), ++ =

+++ = (5.000 - 10.000 /ml)

(ml). (> 10000

analyze the difference between various pre-treatments. A value of P = 0.05 was considered statistically significant.

4.3. Determination of sporulation level

Ten days after the treatment, the effect of essential oils on *C. acutatum* sporulation levels was determined. Determination of sporulation levels was performed using a Thom haemocytometer.

For this purpose, a spore suspension was prepared by adding 5 mL of distilled water to the Petri dish with a culture of isolate C.A.2 (*C. acutatum* J.H. Simmonds). The plate was then viewed under a microscope (Vasic, 2007). The sporulation level is expressed according to the scale by Quesada and Lopez, 1980, where: + = poor sporulation (<5,000 spores/ml), ++ = medium sporulation (5,000 - 10,000 spores/ml) and +++ = abundant sporulation (> 10,000 spores/ml). The experiment was set in four repetitions.

1.

GC/MS

67

99.06%

S. sclarea,

HP-5 MS

1,

(43.17%)

(15.8%).

(9.73%),

(1.43%). EO

(5.13%)

(66.65%),

24.51%,

2.43%,

RESULTS AND DISCUSSION

1. Chemical composition of the essential oil

GC/MS analyses revealed the presence of 67 constituents making up to 99.06 % of the of the analyzed *S. sclarea* essential oil, and the components identified are listed in order of elution from the HP-5 MS column in the Table 1, along with their retention indices, quantitative data and identification method. The most abundant constituents were monoterpenic ester linalyl acetate (43.17%) and its corresponding alcohol linalool (15.8%). The other principal constituents were identified as germacrene D (9.73%), caryophyllene (5.13%) and sclareol (1.43%).

The EO was dominated by the oxygenated monoterpenes (66.65%) followed by the sesquiterpene hydrocarbons amounted to 24.51%, oxygenated sesquiterpenes

2.05%	1.71%	2.43%, monoterpenes 2.05% and diterpenes 1.71%. The mechanisms of EO action against plant pathogens are still not well known. It has been suggested that biophysical, and the intensity of biological, characteristics of essential oils depends on chemical structure of their components.
		Along with the synergy, antagonism or additive effects of the EOs constituents, the influence on the targeted pathogen, structural and morphological changes, or perhaps a specific membrane interaction are also worth being considered. For instance, terpenes comprising the vast majority of the essential oils are responsible for the hydrophobic characteristics of EOs allowing their diffusion through the fungal membrane, affecting intracellular metabolic pathways and organelles.
		Obvious harmful effects on the morphology of cell membranes, release of intracellular components, small ions such as potassium and phosphate, followed by macromolecular substances such as DNA and others were observed in antimicrobial mechanism of <i>S. sclarea</i> essential oil action (Cui et al., 2015).
2015). al. 2019,	<i>S. sclarea</i> (Cui et al., Kumar et	Recent study by Kumar et al. 2019, reported fungitoxicity of Clary Sage essential oil, alone, and in combination with linalyl acetate and linalool, against <i>Aspergillus nidulans</i> (86.37%) and <i>Alternaria alternata</i> (98.37%).
<i>Aspergillus nidulans</i> (86.37%) <i>Alternaria alternata</i> (98.37%).	<i>Aspergillus</i> <i>Alternaria alternata</i>	Reduction of ergosterol, one of the principal sterol providing rigidity and structural integrity to fungal plasma membrane, and prominent leakage of vital ions (Ca^{2+} , K^+ and Mg^{2+}) and UV-absorbing materials in a dose dependent manner is proven (Kumar et al., 2019). Abovementioned demonstrate the complexity of the structural changes underlying the antifungal mechanism induced by <i>S. sclarea</i> essential oil in crop pathogens.
UV	(Ca^{2+} , K^+ Mg^{2+}) (Kumar et al., 2019).	
<i>S. sclarea</i>		

sclarea

Table 1. Chemical composition of the aerial parts essential oil from *Salvia sclarea*

Rt	RI ^a	Compound	Content ^b [%]	Compound class	Identification method ^d
4.44	932	-Pinene	0.11	MT	MS, RI, Col
4.71	946	Camphene	0.02	MT	MS, RI, Col
5.09	969	Sabinene	0.02	MT	MS, RI, Col
5.18	974	-Pinene	0.11	MT	MS, RI, Col
5.32	988	-Myrcene	0.73	MT	MS, RI, Col
6.05	1020	<i>p</i> -Cymene	0.02	MT	MS, RI, Col
6.14	1024	D-Limonene	0.27	MT	MS, RI, Col
6.23	1032	(<i>Z</i>)- -Ocimene	0.28	MT	MS, RI
6.47	1044	(<i>E</i>)- -Ocimen	0.44	MT	MS, RI
7.05	1067	<i>cis</i> -Linalool oxide	0.04	MT	MS, RI, Col
7.40	1084	<i>trans</i> -Linalool oxide	0.05	MT	MS, RI, Col
7.44	1086	Terpinolene	0.1	MT	MS, RI
7.69	1095	Linalool	15.80	MT	MS, RI, Col
8.98	1154	Nerol oxide	0.01	MT	MS, RI
9.33	1165	Borneol	0.05	MT	MS, RI
9.61	1174	Terpinen-4-ol	0.04	MT	MS, RI, Col
9.79	1179	<i>p</i> -Cymen-8-ol	Tr	MT	MS, RI
9.92	1186	-Terpineol	3.20	MT	MS, RI, Col
10.06	1200	Dodecane	0.154	O	MS, RI
10.48	1214	Linalool formate	0.08	MT	MS, RI
10.84	1227	Nerol	0.57	MT	MS, RI, Col
11.16	1235	Neral	0.04	MT	MS, RI, Col
11.58	1254	Linalool acetate	43.13	MT*	MS, RI, Col
11.84	1256	Geraniol	0.05	MT*	MS, RI, Col
12.28	1287	Bornyl acetate	0.08	MT*	MS, RI, Col
12.66	1298	Geranylformate	0.05	MT*	MS, RI, Col
12.59	1300	Tridecane	0.03	O	MS, RI
13.98	1345	-Cubebene	0.12	MT*	MS, RI
13.88	1346	-Terpinyl acetate	0.11	MT*	MS, RI, Col
14.16	1359	Neryl acetate	1.70	MT*	MS, RI, Col
14.64	1379	Geranyl acetate	1.50	MT*	MS, RI, Col
14.68	1374	-Copaene	3.39	ST	MS, RI,
14.92	1387	-Bourbonene	0.54	ST	MS, RI,
15.00	1390	-Cubebene	1.07	ST	MS, RI
15.81	1417	(<i>E</i>)-Caryophyllene	5.13	ST	MS, RI, Col
16.03	1428	-copaene	0.15	ST	MS, RI
16.09	1432	- <i>trans</i> -Bergamotene	0.06	ST	MS, RI
16.38	1451	Isogermacrene D	0.05	ST	MS, RI
16.62	1452	-Humulene	0.36	ST	MS, RI, Col
16.83	1458	<i>allo</i> -Aromadendrene	0.07	ST	MS, RI
17.14	1478	-Muurolene	0.17	ST	MS, RI
17.31	1484	Germacrene-D	9.73	ST	MS, RI
17.42	1489	Eremophilene	0.54	ST	MS, RI
17.66	1500	Bicyclogermacrene-2	1.82	ST	MS, RI
17.72	1500	-Muurolene	Tr	ST	MS, RI
18.05	1513	-Cadinene	0.11	ST	MS, RI
18.23	1522	-Cadinene	0.82	ST	MS, RI
18.79	1544	-Calacorene	0.04	ST	MS, RI
19.05	1549	Salviadienol	0.05	ST	MS, RI
19.32	-	1,5-epoxysalvia-4(14)-ene	0.12	ST	MS, RI
19.58	1577	Spathulenol	0.46	ST	MS, RI
19.69	1582	Caryophyllene oxide	0.85	ST	MS, RI
19.96	1595	4(14)-Salvia-1-one	0.46	ST	MS, RI

20.17	1639	<i>allo</i> -Aromadendrene epoxide	0.08	ST*	MS, RI
20.39	-	Torilenol	0.13	ST*	MS, RI
20.98	1640	Isospathulenol	0.15	ST*	MS, RI
21.27	1649	-Eudesmol	0.09	ST*	MS, RI
21.35	-	Cadina-1(10),4-dien-8-ol	0.20	ST*	MS, RI
21.92	-	Caryophylla-3(15),7(14)-dien-6-ol	0.08	ST*	MS, RI
22.06	1690	Eudesma-4(15),7-dien-1-ol	0.06	ST*	MS, RI
26.20	1906	Sclareoloxide	0.48	O	MS, RI
27.01	1908	Isopimara-8,15-diene	0.06	DT	MS, RI
27.72	1949	geranyl-terpinene	0.06	DT	MS, RI
28.28	1994	Manool oxide	0.04	DT	MS, RI
29.46	-	9-(epi)-Sclarene	0.06	DT	MS, RI
29.48	2055	Manool	0.06	DT	MS, RI
32.38	2222	Sclareol	1.43	DT	MS, RI
Total identified (%)		97.85			
/ Number of components: 67					
/ Monoterpenes 2.05%					
/ Oxygenated monoterpenes 66.65%					
/ Sesquiterpenes 24.51%					
/ Oxygenated sesquiterpenes 2.43%					
/ Diterpenes 1.71%					
/ Others 0.5 %					
) (RI) DB-5MS C7 – C32					
n- . b) ; tr, (<0,02%). c) , Col,					
: RI, ; MS,					
a) Linear retention indices (RI) determined experimentally on the DB-5MS column relative to a series of C7 – C32 n-alkanes.					
b) Values are means of the individual analysis; tr, trace amounts (< 0.02%).					
c) Compound identification: RI, retention indices matching with literature data; MS, mass spectra matching, Col, coinjection with pure reference compound					

Džami et al. reported the chemical composition of *S. sclarea* (Džami et al., 2008). The amount of linalyl acetate (52.83%) and linalool (18.18%) were comparable high, as are in our material, while the percentage of the other components did not display many differences in content, but exhibited quantitative variation. Namely, -terpineol (5%), -pinene (4.57%), limonene (1.55%) and -caryophyllene (1.83%) were one of the, in total of 34, dominant identified compounds, while in our sample they represented just the minor portion of the analyzed EO. The same authors group accessed the antifungal activity evaluation against several micromycete species. A concentration of 25 µL/ml showed minimal fungicidal concentration (MFC) against *Aspergillus*, *Penicillium*, *Fusarium* sp., *Trichoderma viride*, MFC *M. mucedo*

Džami et al. have previously reported the chemical composition of *S. sclarea* EO originating from Serbia (Džami et al., 2008). According to their study, the amount of linalyl acetate (52.83%) and linalool (18.18%) were comparable high, as are in our material, while the percentage of the other components did not display many differences in content, but exhibited quantitative variation. Namely, -terpineol (5%), -pinene (4.57%), limonene (1.55%) and -caryophyllene (1.83%) were one of the, in total of 34, dominant identified compounds, while in our sample they represented just the minor portion of the analyzed EO. The same authors group accessed the antifungal activity evaluation against several micromycete species. A concentration of 25 µL/ml of the EO showed minimal fungicidal concentration (MFC) against *Aspergillus*, *Penicillium*, and *Fusarium* sp., and *Trichoderma viride*, while the MFC values for *M. mucedo* and *A. viride*, and *C. albicans*

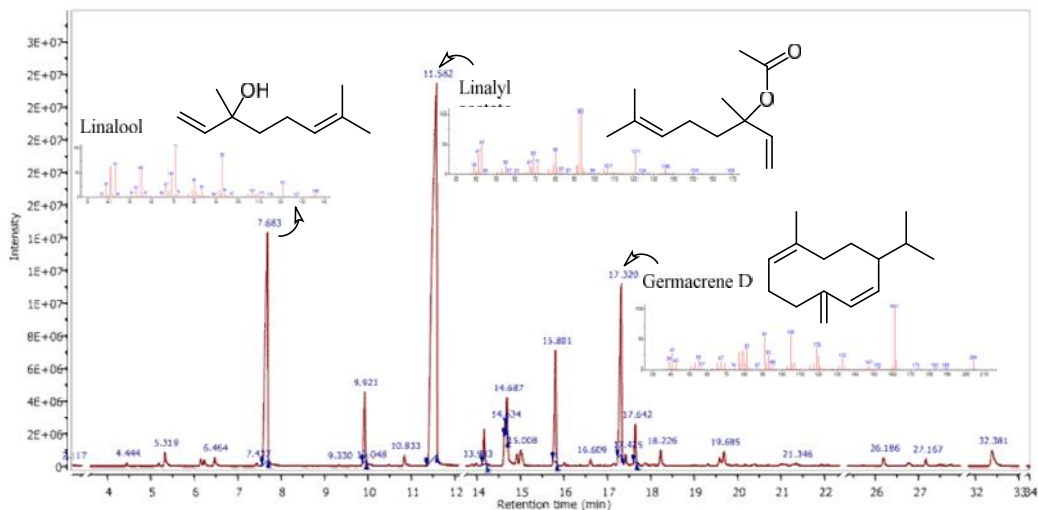
<i>A. viride</i> <i>C. albicans</i> 15 µL/mL 10 µL/mL,	were 15µL/mL and 10 µL/mL, respectively. Fungistatic and fungicidal activities of the oil against darkly
<i>cladosporioides</i> ,	<i>C. cladosporioides</i> attacking both the leaves and fruits of
<i>T. grophytes</i> , 2,5 µL/ml 5 µL/ml.	many plants and <i>T. grophytes</i> were at the concentrations of 2.5 µL/mL and 5 µL/mL.
<i>A.alternata</i> , <i>P. helianthi</i> <i>C. fulvum</i> ,	In the case of <i>C. fulvum</i> , <i>A.alternata</i> , <i>P. helianthi</i> and <i>P. macdonaldii</i> , the minimal
<i>P. macdonaldii</i> (MLC) 2,5 µL/ml (Džami et al., 2008).	lethal concentration (MLC) was proven to be 2.5 µL/mL (Džami et al., 2008).
<i>acutaum</i>	Encouraged by their results we decided to estimate the antifungal potential of our Clary Sage EO against another very important fungal pathogen, <i>C. acutaum</i> and possibly correlate the EO constituents content with the exhibited antifungal activity.
<i>S. sclarea</i>	The presence of linalyl acetate and linalool as the major <i>S. sclarea</i> EO secondary metabolites corroborate to the results presented by Pitarokili and Kuzma (Pitarokili et al., 2002; Kuzma et al., 2009). Pitarokili et al. pointed on the dose-dependent inhibition of fungal soil-borne pathogens growth caused by the Clary Sage EO treatment. Inhibition of <i>S. sclerotiorum</i> mycelia radial growth (1000 µL/L), <i>S.cepivorum</i> (2000 µL/L) and <i>F. oxysporum</i> f. sp. <i>dianthi</i> (2000 µL/L) together with fungicidal effect on <i>S. sclerotiorum</i> at the concentration of 2000 µL/L were recorded.
Pitarokili Kuzma (Pitarokili et al., 2002; Kuzma et al., 2009). Pitarokili et al.	The corresponding EC50 values of the essential oil of <i>S. sclarea</i> obtained for each fungus (<i>S. sclerotiorum</i> (EC50) 492.55 µL/L; <i>S. cepivorum</i> (EC50) 544.17 µL/L; and <i>F. oxysporum</i> f. sp. <i>dianthi</i> (EC50) 584.36µL/L) are presented in the same work. Pure commercial linalool and linalyl acetate were also tested, independently, for their inhibitory action on the radial growth of the same pathogens, and the various degrees of inhibition depending on the fungi tested were recorded (Pitarokili et al., 2002). Having the all above mentioned in mind it
<i>S. sclerotiorum</i> (1000 µL/L), <i>S.cepivorum</i> (2000 µL/L) <i>F. oxysporum</i> f. sp. <i>dianthi</i> (2000 µL/L)	
<i>S. sclerotiorum</i> 2000 µL/L.	
EC50	
<i>S. sclarea</i> , (<i>S. sclerotiorum</i> (EC50) 492,55 µL/L; <i>S. cepivorum</i> (EC50) 544,17 µL/L; <i>F. oxysporum</i> f. sp. <i>dianthi</i> (EC50) 584.36µL/L)	
e	
(Pitarokili et al., 2002).	

S. sclarea (47.4%), (12.7%) (22.1%)
 23
albicans.
 2 mg/ml, MIC (>2 mg/mL),
S. sclarea (49.02%), (19,2%) (Moretti et al., 1997).
S. sclarea (32.97%), (16.85%), (7,57%) (Torres et al., 1997).
 Carruba et al. D (10.56%), (9.73%), (Carruba et al., 2002) (1).
S. sclarea, (81.4%) (10.7%), IC₅₀, (HL-60, K562) (MCF-7)

might be possible that linalol influenced the inhibition of our tested pathogen growth, also. Considering the results, regarding the Sardinian samples of *S. sclarea*, in which -terpineol (47.4%), -terpinyl acetate (22.1%) and linalil acetate (12.7%) were the dominant ones of in total 23 identified constituents, significant microbiostatic action against fungus *C. albicans* was stated. -terpineol, present in our chromatogram also, tested alone, showed a candidicidal activity at doses higher than 2 mg/mL, while linalool exhibited weaker MIC values (>2mg/mL) pointing on the synergistic action of the single components, constituents of the EO.

Major constituents of the analyzed essential oil of Sardinian *S. sclarea* were very distinctive from our sample, dominating methyl chavicol (49.02%), never reported before from this species, and linalyl acetate (19.2%) (Moretti et al., 1997). Spanish specimens of *S. sclarea* prevailed by linalool (32.97%), linalyl acetate (16.85%), germacrene D (7.57%) and terpinen-4-ol (5.63%) (Torres et al., 1997). Investigating the Italian biotype of Clary Sage Carruba et al. pointed on the significance of the high content of germacrene D (10.56%), also present in our sample in the high yield (9.73%), in the possible exploiting of this species essential oil in plant protection (Carruba et al., 2002) (Figure 1).

French specimens of *S. sclarea*, yielding the high percentage of linalyl acetate (81.4%) and linalool (10.7%), showed activity equivalent to IC₅₀ values obtained with doxorubicin against cell suspensions of selected hemopoeitic tumors (HL-60, K562) and solid tumors (MCF-7), and because of the complexity oil composition it was not easily explainable which constituent was the active one.



. 1.

S. sclarea,

Fig. 1. Total ion current chromatogram of the essential oil of *S. sclarea*, with the mass spectrums and chemical formulas of dominant oil constituents

(Foray et al., 1999).

, P-388, KB NSCLC-N6, 0.06% (Džami et al., 1998; Džami et al., 2008).

1.43% (Dimas et al., 1998; Džami et al., 2008).

(Bailey et al., 1975)

(Ulubelen et al., 1994).

(Elnir et al., 1991), (Moretti et al.,

Of course, a synergistic effect of some compounds in the secondary metabolites mixture raises as a reasonable assumption. (Foray et al., 1999). Sclareol, a labdane-type diterpen, with high antimicrobial activity and dose, and time dependent cytostatic and cytotoxic potential against a panel of human leukemic cell lines, P-388, KB and NSCLC-N6 cell lines was present in only 0.06 % in Džami et al. plant material, while in our sample it comprised 1.43% of the oil (Dimas et al., 1998; Džami et al., 2008). This bioactive diterpene induced the reduction of the severity of rust infection in French beans and wheat (Bailey et al., 1975), and the inhibition of the radial extension of fungal colonies grown in agar (Ulubelen et al., 1994). It can be concluded that different Clary Sage chemotypes have been identified. Besides the most common linalyl acetate/linalool dominant, -terpineol dominant, geraniol/geranyl acetate-rich chemotype (Elnir et al., 1991), a methyl chavicol-rich chemotype (Moretti et al., 1997), a germacrene-D-rich chemotype (Carruba et al., 2002), and very recently,

1997), (Carruba et al., 2002), -	D	-thujone, thujene, and manool oxide/phytolchemotypes from Tunisia (Taarit et al., 2011).
	/	
	(Taarit et al., 2011).	
	GC-MS	According to our extensive GC-MS analysis
	<i>S. sclarea</i>	the <i>S. sclarea</i> essential oil from south- eastern Serbia in our hands is rich in linaly lacette, linalool, but also contains high percentage of germacrene D (Figure 1).
(1).	D	
	-	Eugenol represents the dominant constituent of the <i>S. aromaticum</i> (L.) Merr. & L.M. Perry and <i>C. verum</i> J.Presl EOs, while carvacrol, another aromatic compound with the phenolic OH group present in its structure, from <i>O. vulgare</i> L. demonstrate antifungal activity due to the hydrogen bond formation with the targeted enzyme active cite (Farag et al., 1989).
Merr. & LM Perry	<i>S. aromaticum</i> (L.)	
	<i>C. verum</i> J.Presl EOs,	
	OH	
<i>vulgare</i> L.	, O.	
	-	
	-	
	(Farag et al., 1989).	
	-	EO tested in this work does not contain these particular oxygenated monoterpenes, but it does include compounds with a similar chemical structure, so it could be speculated that the inhibition of mycelial growth and an alteration of the sporulation level of <i>C. acutatum</i> is accomplished partly due to their presence.
	(66.65%),	
	,	
	,	
	<i>C. acutatum</i>	
	,	
	,	
<i>flavus</i>	,	
	A.	Aliphatic alcohols, geraniol, nerol and citronellol, some of which present in our sample, suppressed the growth of <i>A.</i> <i>flavus</i> and consequently prevented the formation of aflatoxin, (Mahmound, 1994) and EOs containing aliphatic alcohols and phenols exhibited significant action against <i>A. aegyptiaceus</i> , <i>P. cyclopium</i> and <i>T. viride</i> (Megalla et al., 1980).
1994),	(Mahmound,	
	,	
	.	
<i>aegyptiaceus</i> , <i>P. cyclopium</i>	<i>T. viride</i>	
(Megalla et al., 1980).		
	,	
	,	
	.	
	,	
	C.	Above-mentioned results, together with our report suggest that aliphatic alcohols could have antifungal action against a broad spectrum of fungi. Due to the complexity of the EO composition in our hands, it was not straightforward to deduce which is the active one attributed for the antifungal activity against <i>C.</i> <i>acutatum</i> , pointing on the possible synergistic effect of a number of compounds in the mixture.
<i>acutatum</i> .		

2.
2.1.

Colletotrichum acutatum

S. sclarea

C.

0.04 µL/ml

2) 0.04 µL/ml

0.8 µL/ml

(MLC)

2. Antifungal activity
2.1. Effect of volatile phase of essential oils on the growth of *Colletotrichum acutatum* isolates

S. sclarea essential oil inhibited the growth of *C. acutatum* mycelium and the percentage of the inhibition was concentration dependent. To be exact, inhibition of mycelial growth was observed at concentrations of 0.04 µL/mL of air and higher, considering the essential oil of Clary Sage. The minimum inhibitory concentration (MIC) of the tested EO (Table 2; Figure 2) was at the concentration of 0.04 µL/mL of air, while the minimal lethal concentration (MLC) was at the concentration of 0.8 µL/mL of air.

2.
C. acutatum

Table 2. The effect of volatile phase of the essential oil on the growth of *C. acutatum* mycelia

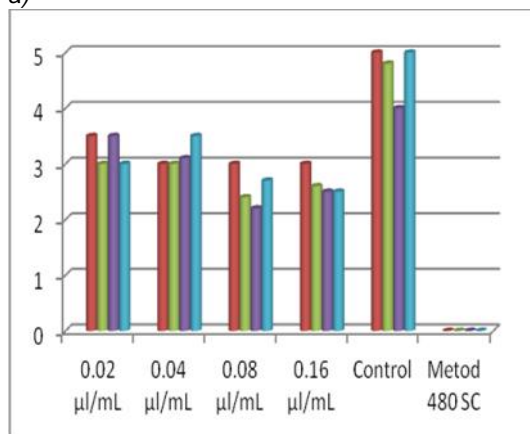
p.s. con	0.02 (µl/mL)	0.04 (µl/mL)	0.08 (µl/mL)	0.16 (µl/mL)	Control	Metod 480 SC
<i>Salvia sclarea</i> L.	3.25 ^b	3.15 ^b	2.58 ^c	2.65 ^c	4.70 ^a	-

* (P = 0,05)

(p = 0,05)

* (P=0.05) The data in rows marked by the same letter are not statistically significantly different based on Duncan test (p=0.05)

a)



b)



Salvia sclarea

, b)

(*Fragaria x ananassa* Olympus)

. 2.)

Colletotrichum acutatum in vitro
Colletotrichum acutatum

Duchesne ex Rozier [chiloensis x virginiana]
CX31 c)

Fig. 2. a) The effect of volatile phase of the *Salvia sclarea* essential oil on the growth of *Colletotrichum acutatum* isolate *in vitro* after the seven-day exposure, b) Asexual spores of the fungus *Colletotrichum acutatum* with measurement scale on the strawberry (*Fragaria x ananassa* Duchesne ex Rozier [chiloensis x virginiana]) host from Serbia under the Olympus CX31 microscope c) Symptoms of strawberry anthracnose on fruits

2.2.

C.A.2 (*C. acutatum*)
 -
 (2).
S. sclarea,
 ,
C. acutatum *in vitro* .
 0.08 µL/mL.
 0.02-0.08 µL/ml
C. acutatum.
 (MLC)
C. acutatum
 . Vasi , 2007 ,
 (Vasi , 2007).
 (Metod 480 SC),
 ,
C. acutatum,
 ,
 (3).

3.

C. acutatum

Table 3. Influence of the *Salvia sclarea* essential oil on the sporulation level of *C. acutatum*

Sporulation level	0.02 (µl/mL)	0.04 (µl/mL)	0.08 (µl/mL)	0.16 (µl/mL)	Control	Metod 480 SC
<i>Salvia sclarea</i>	++	++	++	++	+++	-

in vitro

2.2. Determination of sporulation level

- EO greatly influenced the sporulation ability of the C.A.2 (*C. acutatum*) test isolate to form conidia in larger or smaller numbers (Table 2).
- Thus, the EO of *S. sclarea*, compared to the control, has shown a medium sporulation level for all of the applied concentrations, and an inhibitory effect on the growth of *C. acutatum* mycelium under *in vitro* conditions. The essential oil exhibited a strong inhibitory effect at concentrations higher than 0.08 µL/mL. The minimum inhibitory concentrations of Clary Sage oil was 0.02-0.08µL/mL of air, for the tested *C. acutatum* isolate.
- The minimum lethal concentration (MLC) was expressed after five days. The sporulation level of the tested *C. acutatum* isolate was different depending on the applied concentration. Vasi , 2007 states that the level of sporulation significantly affects the speed and the intensity of infection (Vasi , 2007).
- The referent antimycotic captan (Metod 480 SC), phtalimide class fungicide used as a positive control, manifests its fungicidal activity by stopping the growth of the mycelium of the *C. acutatum* making a protective barrier on the leaves and fruits preventing the entrance of pathogen and blocking the energy production, was being used (Table 3).

Salvia sclarea

- Numerous *in vitro* studies demonstrate the high efficacy of essential oils against bacteria and fungi contaminants.

	. Beatovi et al.	Beatovi et al. studied the antifungal activity of varying concentrations of <i>O. sanctum</i> L. EO on diverse fungal pathogens: <i>A. ochraceus</i> , <i>A. versicolor</i> , <i>A. niger</i> , <i>A. fumigatus</i> , <i>T. viride</i> and <i>P. funiculosum</i> (Beatovi et al., 2013). Antifungal activity of 18 essential oils was evaluated against <i>V. fungicola</i> var. <i>fungicola</i> (Preuss) Hassebrauk, <i>M. pernicioso</i> (Magnus) Delacroix <i>Cladobotryum</i> spp. (Cooke) <i>A. bisporus</i> (Lange) Imbach., <i>in vitro</i> .
<i>sanctum</i> L.	O.	
	: <i>A. ochraceus</i> , <i>A. versicolor</i> , <i>A. niger</i> , <i>A. fumigatus</i> , <i>T. viride</i> <i>P. funiculosum</i> (Beatovi et al., 2013).	
	18	
	<i>V. fungicola</i> var. <i>fungicola</i> (Preuss) Hassebrauk, <i>M. pernicioso</i> (Magnus) Delacroix <i>Cladobotryum</i> spp. (Cooke) <i>A. bisporus</i> (Lange) Imbach., <i>in vitro</i> .	
	-	Of the essential oils analyzed, cinnamon, clove, thyme, and tea tree showed the strongest antifungal activity against all investigated mycopathogens, with Minimum Fungicidal Concentration (MFC) being 0.02 µL/mL of air for isolates tested (Tanovi et al., 2006).
	-	
	-	
	-	
	(MFC) 0.02 µL/ml (Tanovi	
at al., 2006). Grahovac et al.		Grahovac et al. tested 56 essential oils against the anthracnose causative pathogens <i>C. acutatum</i> and <i>C. gloeosporioides</i> , of which oregano and thyme oils have shown the highest antifungal activity. The lowest lethal concentration of oregano oil was at the 0.02 µL/mL of air for the <i>C. acutatum</i> isolate (Grahovac et al., 2014).
56	-	
	<i>C. acutatum</i> C.	
<i>gloeosporioides</i> ,	-	
	-	
	0.02 µL/ml	
	<i>C. acutatum</i> (Grahovac	
et al., 2014).	Duduk et al.	
(2010)	<i>in vitro</i>	
	-	In addition, Duduk et al. (2010) studied the antifungal effect of three essential oils extracts: <i>T. vulgaris</i> , <i>C. zeylanicum</i> and <i>S. aromaticum</i> on <i>C. acutatum</i> , <i>in vitro</i> . All of the tested essential oils inhibited the growth of <i>C. acutatum</i> mycelium at the concentrations of 46 µL/L of air and higher, and the percentage of inhibition was dependent on the amount of essential oil applied. Daferera et al. examined the antifungal activity of eight essential oils, including one <i>Salvia</i> sp., against <i>B. cinerea</i> , <i>Fusarium</i> spp. and <i>C. michiganensis</i> subsp. <i>Michiganensis</i> on an artificial growth media (Daferera et al., 2003). The growth of the above mentioned pathogens was completely inhibited by oregano (thymol dominant, 63.7%), thyme, dictamnus, and marjoram (all rich
	: <i>T. vulgaris</i> , <i>C. zeylanicum</i> S.	
<i>aromaticum</i>	<i>C. acutatum</i> .	
	<i>C. acutatum</i>	
	46 µL/L	
	-	
	. Daferera et al.	
<i>Salvia</i> sp., <i>B. cinerea</i> ,		
<i>Fusarium</i> spp. <i>C. michiganensis</i> subsp.		
<i>michiganensis</i>	<i>Michiganensis</i>	
	(Daferera et al., 2003).	
	-	
	(

63.7%),
()
300 µg/ml),
(),
(
31.5% 59.5%)
-
et al.

. Velluti
1 ()
,
)
F. proliferatum,

proliferatum
et al., 2003).

1 *F.*
(Velluti

in carvacrol) essential oils at relatively low concentrations (85-300 µg/mL), while the essential oils of lavender (linalool and linalyl acetate dominant), rosemary and sage (both eucalyptol dominant with 31.5% and 59.5%, respectively) exhibited less inhibitory activity.

Velluti et al. examined the effect of five essential oils on growth and fumosin B1 (hepatotoxic and nephrotoxic toxin occurring mainly in corn, wheat and other cereals) production by three different isolates of *F. proliferatum*, suggesting the synergistic action of the dominant and the minor compounds of the EO tested.

The same authors group found that the cinnamon and oregano essential oils inhibited the growth of mycelium and reduced the production of fumosin B1 in *F. proliferatum* in corn kernels (Velluti et al., 2003).

Above-mentioned studies, together with many others, suggest that the EOs are a potent source of biopesticides against a great variety of fungal causatives.

CONCLUSIONS

- The control of fungal diseases responsible for significant losses of crop production is considerable problem.
- Persistent use of synthetic pesticides causes pollution enhancement and accordingly, research on and identification of, new non-toxic and non-polluting, active plant constituents, as an alternative natural antifungal biopesticides, for the control of diseases in agriculture is of great importance.

Volatile compounds from plants have antibacterial, antifungal and insecticidal activities, and considered this they can be regarded as a prolific source of natural plant-derived fungicides successfully applied as biocontrol agents in a variety of crops.

<p><i>S. sclarea</i></p> <p><i>in vitro</i></p> <p><i>Colletotrichum acutatum</i> JH Simmonds C.A.2</p> <p><i>S. sclarea</i></p> <p><i>C. acutatum</i>,</p>	<p><i>in</i></p>	<p>The essential oil of <i>S. sclarea</i> is evaluated for its suitability in the <i>in vitro</i> antifungal activity assessment against plant pathogenic fungus <i>Colletotrichum acutatum</i> J.H. Simmonds C.A.2 isolates, for the first time, in order to evaluate other alternative crop protection methods. Clary Sage have previously shown good antifungal activity against several plant pathogens and could possibly serve as a natural alternative to synthetic fungicides for the control of some important fungal diseases.</p> <ul style="list-style-type: none"> - Mechanism of fungicidal and fungistatic action should be yet clarified, and the appropriate approach for the enhanced performance of EO by altering vehicles able to adhere to the effected plant organs, prolonging that way the inhibitory effect on the growth of pathogen, could perhaps be taken into account. <p>All the above-mentioned demand further research in order to evaluate <i>S. sclarea</i> essential oil potential for the purpose of the practical application in the control of the <i>C. acutatum</i>, causative of strawberry anthracnose.</p>
		<p>ACKNOWLEDGEMENTS</p>
		<ul style="list-style-type: none"> - The authors acknowledge the - Ministry of Education, Science and Technological Development of the Republic of Serbia for the financial support (Project 172061).
<p>(172061).</p>		

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